

Sanitation for nurseries



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Why is Sanitation Important?

- Maintaining a clean nursery operation prevents unwanted pests and diseases from entering the nursery
- Sanitation should be considered one of the most important factors in a nursery operation

Sources of inoculum

- People and their vehicles
- Plants
- Dirty pots, tools, and equipment
- Soil
- Water
- Plant debris and cull piles
- Weeds, algae, and associated insects (shoreflies and fungus gnats)

People

- Employee training
- Visitor sign-in at office
- Footbaths at entry to clean areas



1. dirty boots
2. rinsed with water
3. disinfected

Source: ILVO



Remove soil and mud from shoes before using disinfectant footbaths.



1. What kind of access do customers and visitors have to your nursery?

Visitors sign in at office

Only nursery vehicles allowed on property

Loading dock area for deliveries

Public access to retail area, employees only in wholesale and production areas

Full access by customers and their vehicles

Equipment

- Vehicles
- Carts and wheelbarrows
- Hoses
- Tools

P. ramorum positive mud



Delivery vehicles



- Delivery vehicles should be directed to a designated area upon arrival
- Do not allow drivers to sweep out their vehicles on your property
 - Send them away to clean the vehicle before returning
- **You do not know where they have been!**

Keep vehicles on designated roadways

Avoid hitting any puddles in the roadway to
reduce splash on surrounding plant material





Plants

- Incoming plants should be examined on arrival
- Returned plants are potential carriers of disease

Proper plant spacing will reduce stress and disease spread

- Better airflow to reduce humidity
- Plant to plant contact is reduced



Spread of Pr within a nursery

- Very moist conditions needed for infection & disease expression
- Direct contact = very important for plant-to-plant spread
- Aerial dispersal = very unlikely. Spore loads too small?
- Spread via water film and splash dispersal = most likely
- Tipping over in zoospore-containing water film = infection
- -> production nursery: most likely spread via water films

[Within-Field Spread of *Phytophthora ramorum* on Rhododendron in Nursery Settings](#)


Kurt Heungens, Isabelle De Dobbelaere, Bjorn Gehesquière, Annelies Vercauteren, and Martine Maes. In Frankel, Susan J.; Kliejunas, John T.; Palmieri, Katharine M. 2010. Proceedings of the Sudden Oak Death Fourth Science Symposium. Gen. Tech. Rep. PSW-GTR-229. Albany, CA: U.S. Department of Agriculture, Forest Service, Pacific Southwest Research Station. 378 p.

Keep plant debris from accumulating




Pots

- Soil and plant material clinging to used pots and trays are a source of pests, diseases, and weeds
- Pots can be treated with disinfectant, steaming, or hot water dip



Hot water dip tank for 4" pots



This steam cabinet was built on a relatively low budget by a nursery in OR

Disinfectants for tools, pots, and other hard surfaces

- Chlorine bleach – corrosive, do not use on metal tools
- Quaternary ammonium compounds
- Hydrogen peroxide
- Wash organic debris off with water then soak in disinfectant for 10-15 minutes

2. How do you deal with used pots and/or trays at your nursery?

Wash and sanitize prior to re-use

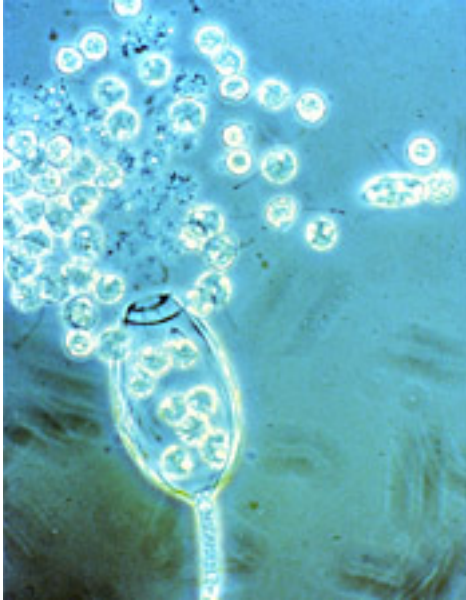
Discard used pots and trays and buy new ones

We re-use dirty pots and trays

Substrate



Diseases that thrive in wet soils



Phytophthora (F.W. Schwenk, Kansas State University)



Plant-parasitic and non-parasitic nematodes recovered from soil by sieving. (Greg Tylka)

- Oomycetes or “water molds”
- Certain fungi
- Bacteria
- Nematodes

Many pathogens produce resistant
“resting spores” that can survive in soil.



Chlamydospores in infected
foliage are a source of Pr
inoculum in soil

The soil phase of *P. ramorum*

- Pr can survive for at least 8 - 11 months in soil or potting media
- Chlamydospores germinate and form sporangia near roots
- Infected root tips seen covered with sporangia
- Pr was detected in root balls of potted Rhododendrons 2 years after purchase



Shishkoff, N. 2007. Persistence of *Phytophthora ramorum* in soil mix and roots of nursery ornamentals. Plant Dis. 91: 1245-1249

Vercauteren et al. 2012. Survival of *P. ramorum* in Rhododendron root balls and in rootless substrates. Plant Pathology
doi:10.1111/j.1365-3059.2012.02627.x

Prevention BMPs for soil:

Cull piles are a breeding ground for diseases. Keep them separate and downhill from production areas.




Soil and media piles should be covered to prevent weeds and pathogens




Media piles should be on an impervious surface with good drainage to prevent contamination, and uphill from production areas.



More prevention BMPs for soil

A photograph showing several black plastic pots containing flowering plants with pink blossoms. The pots are arranged on a green metal bench supported by concrete blocks. The background shows more plants and a wooden structure.

Use pallets or benches
underneath pots

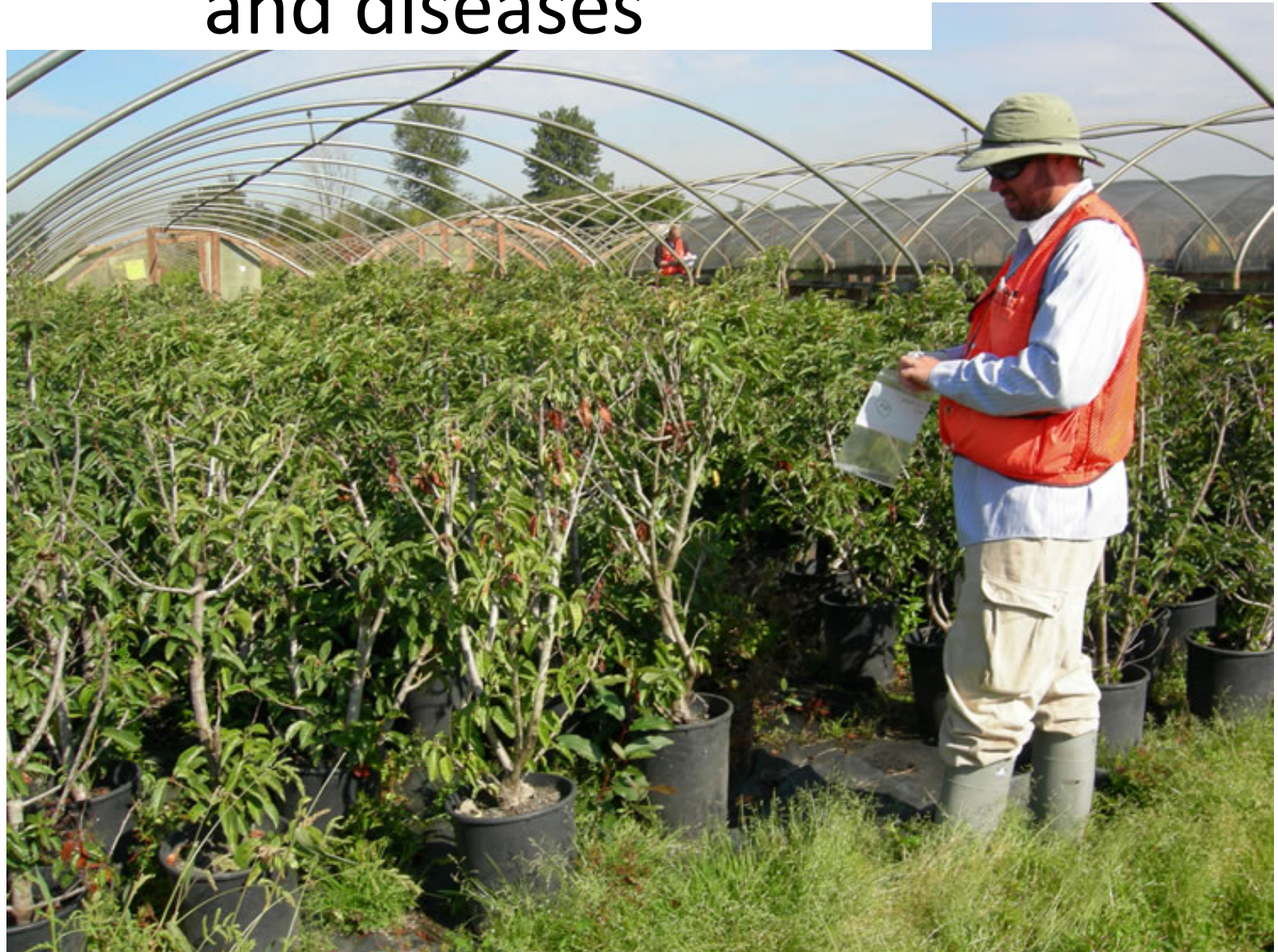
A photograph showing several black plastic pots containing green leafy plants. The pots are placed on a layer of light-colored gravel. The background shows more plants and a wooden structure.

Gravel layer of 4-6" will provide
drainage and separate plants
from soil surface

3. What surface are container plants placed on at your nursery?

Weed mat
Gravel
Bare soil
Pallets
Benches
Other

Weeds are a source of pests and diseases



Algae



Good drainage is important

Fungus gnats

Found in wet soil environments

Feed on algae and fungi.

Larvae feed on plant roots.

Adults can spread plant diseases by moving from plant to plant.



Whitney Cranshaw, Colorado State University, Bugwood.org



University of Nebraska
Department of Entomology

Animal vectors for *Phytophthora*



Fig. 5. Germinated sporangia of *Phytophthora ramorum* in snail frass. Bar = 55 μ m.

Snails – chlamydospores and sporangia

Shore fly larvae – chlamydospores

Shore fly adults – none

Fungus gnat larvae – chlamydospores

Fungus gnat adults - none

Hyder, N., Coffey, M. D., and Stanghellini, M. E. 2009. Viability of oomycete propagules following ingestion and excretion by fungus gnats, shore flies, and snails. *Plant Dis.* 93:720-726.

You might have contaminated soil if

- In-ground plants are symptomatic in an area
- Batches of potted plants from a media pile are symptomatic
- There is standing water indicating a drainage problem
- The soil tests positive for *Phytophthora*



Photo: Jay W. Pscheidt.

Prevent re-contamination of treated soil

Before treating the soil, ensure that the problem doesn't happen again by

- Relocating any nearby cull piles
- Place media on impervious surface with good drainage
- Improve drainage to route water away from plants



Treatments for contaminated soil

Soil amendments/
Biocontrol

Solarization

Steam

Fumigation



Some soil organisms are beneficial

Cellulase producing fungi

Bacteria and actinomycetes
produce antibiotics

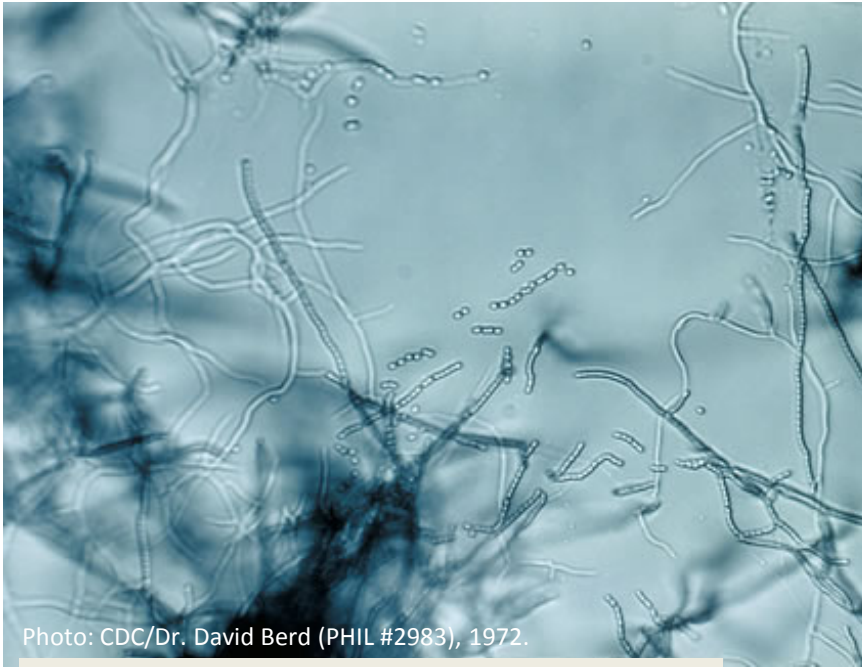


Photo: CDC/Dr. David Berd (PHIL #2983), 1972.

Streptomyces spp. are common in
soil and produce antibiotics



Several species of *Trichoderma* are
antagonistic to *Phytophthora* spp.

Organic soil treatments

Mulches, composts, and bark chips contain antagonistic bacteria and fungi



Photo: Wilson's Garden Center, Utica, OH

Phytophthora + Trichoderma



Normal Pr chlamydospore



Pr chlamydospore parasitized by
Trichoderma

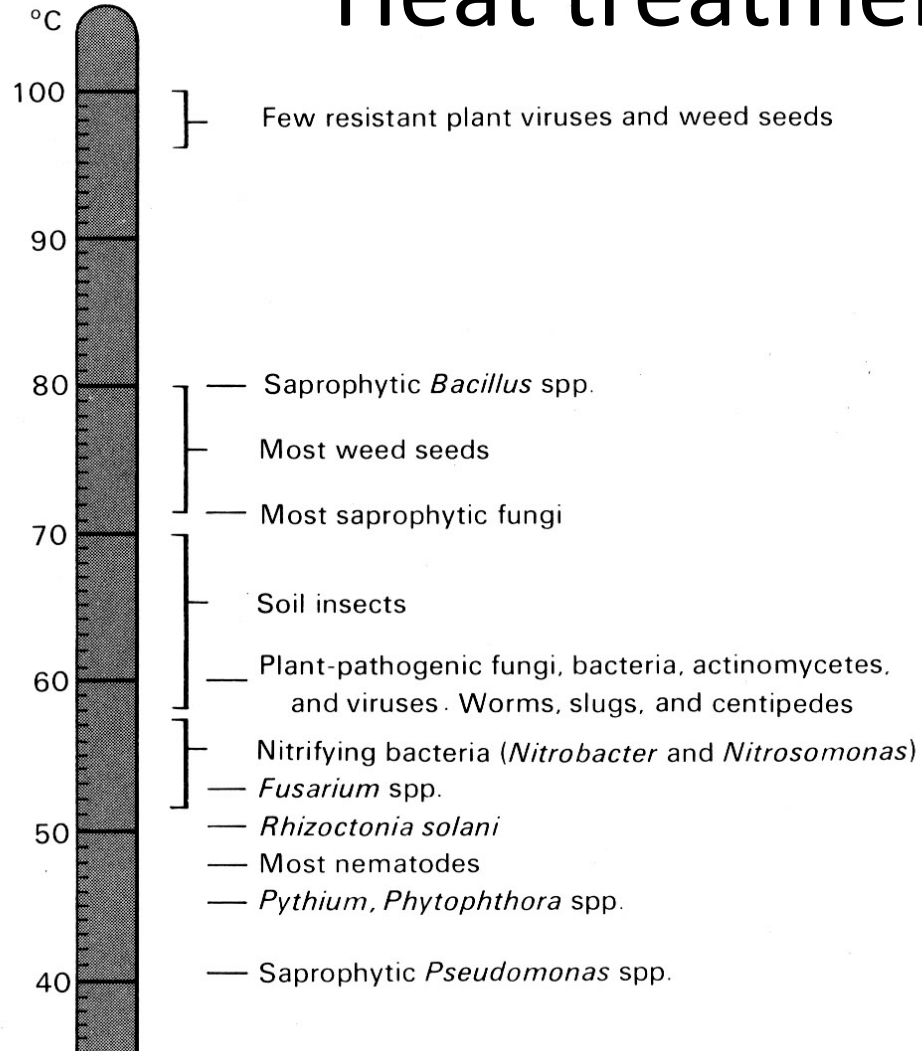
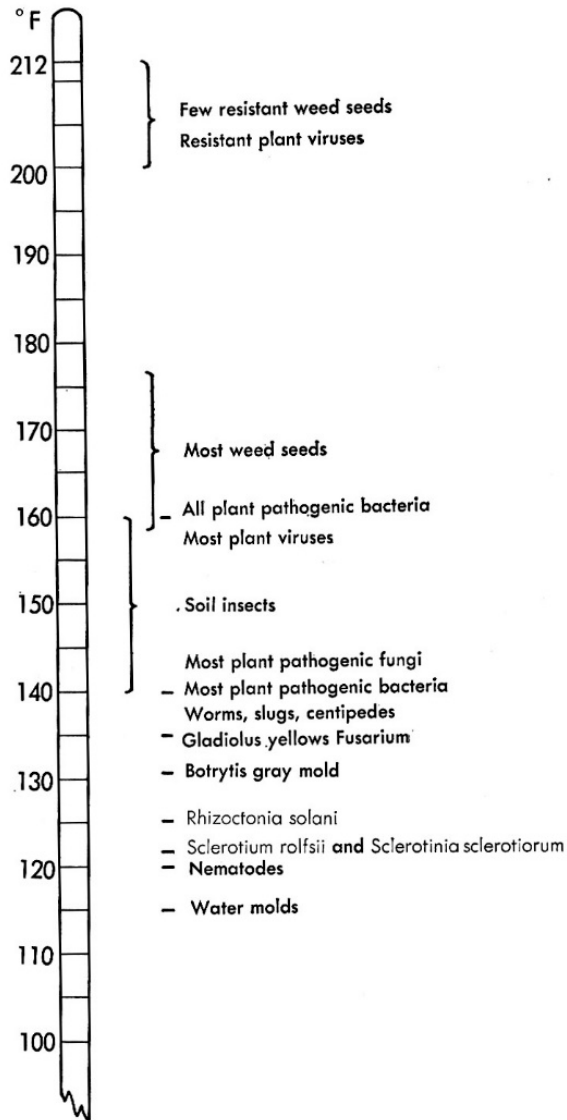
Chemical fumigation

- For large areas of soil
- Kills beneficial organisms
- Dazomet – granular formulation
- Chloropicrin
- Metam sodium – can be worked into soil or applied through irrigation

Check label for approved crops and application methods before use



Heat treatment

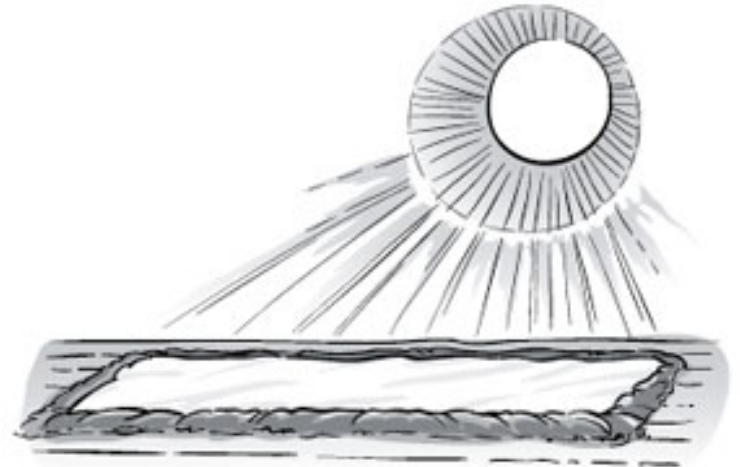
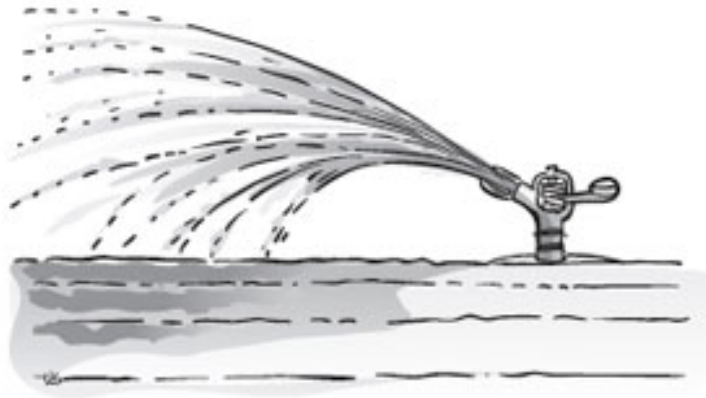
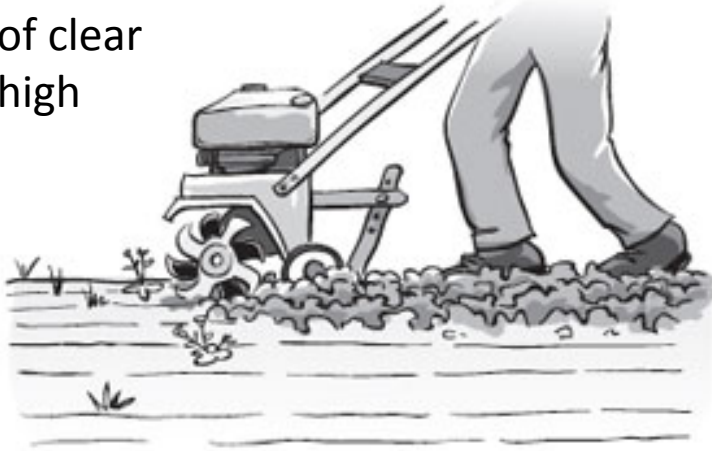


Temperatures necessary to kill various groups of soil organisms

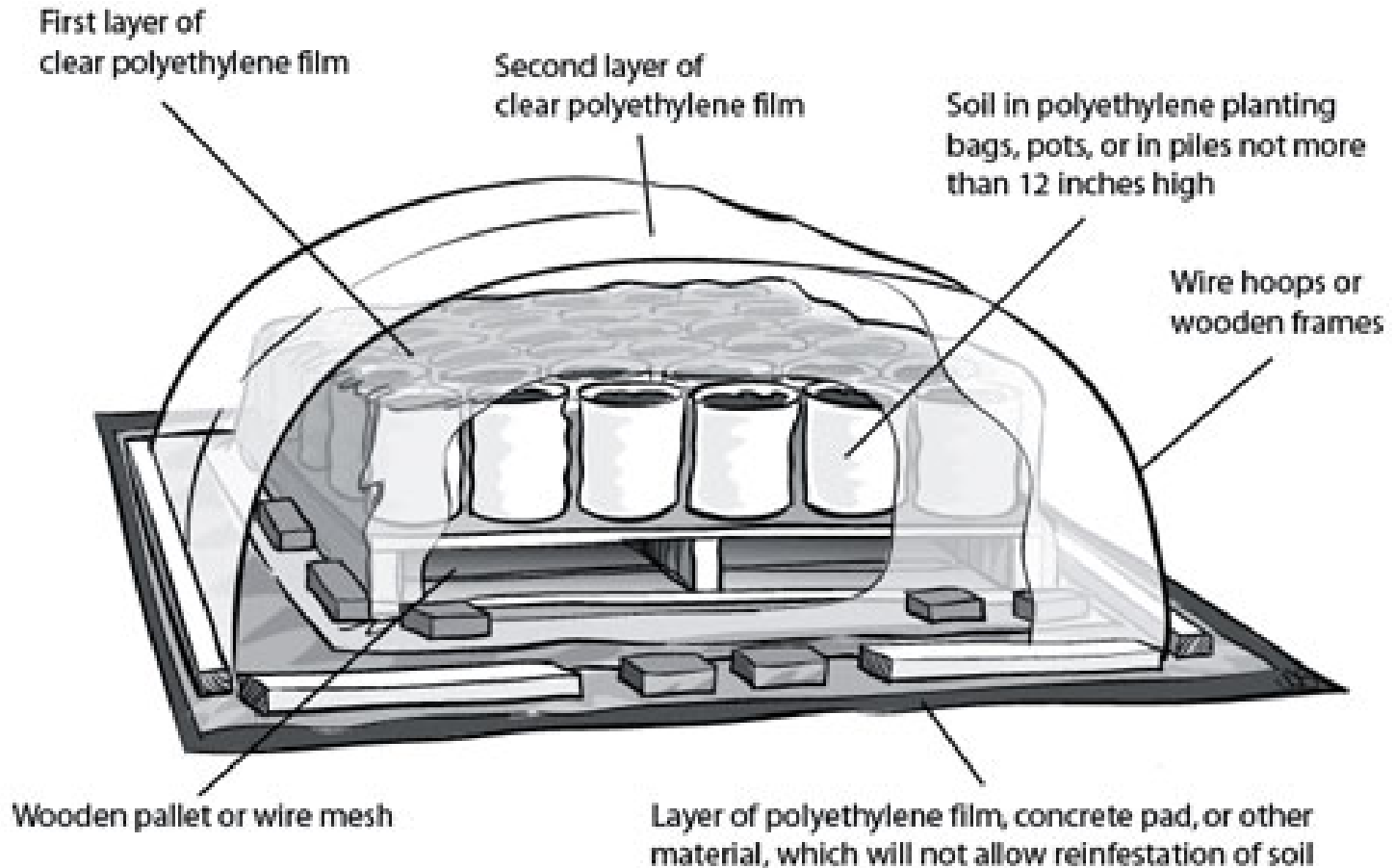
Solarization

Heats top layer of soil to kill pests and pathogens.

Need 4-6 weeks of clear skies, long days, high temperatures.



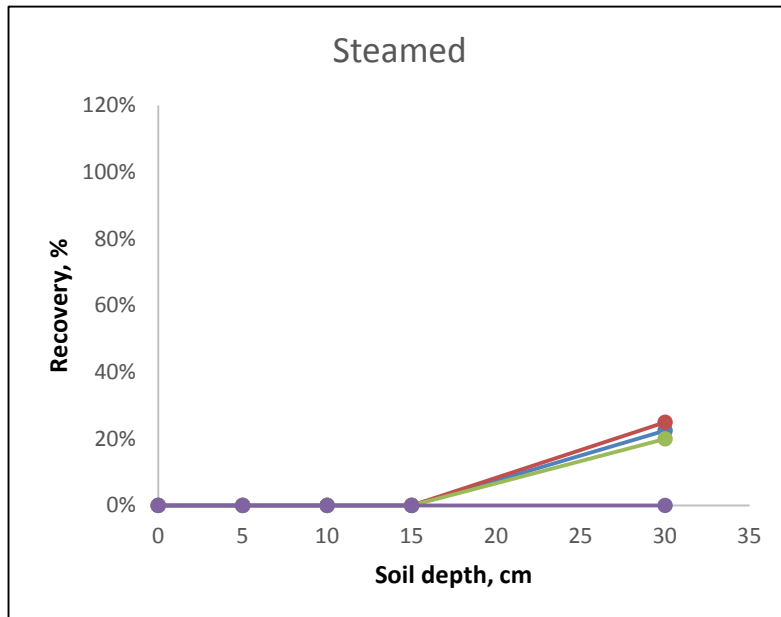
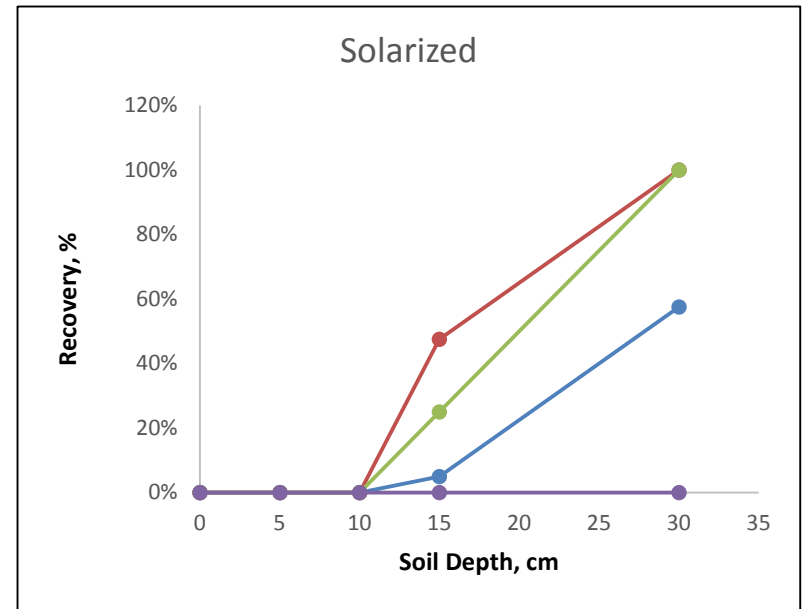
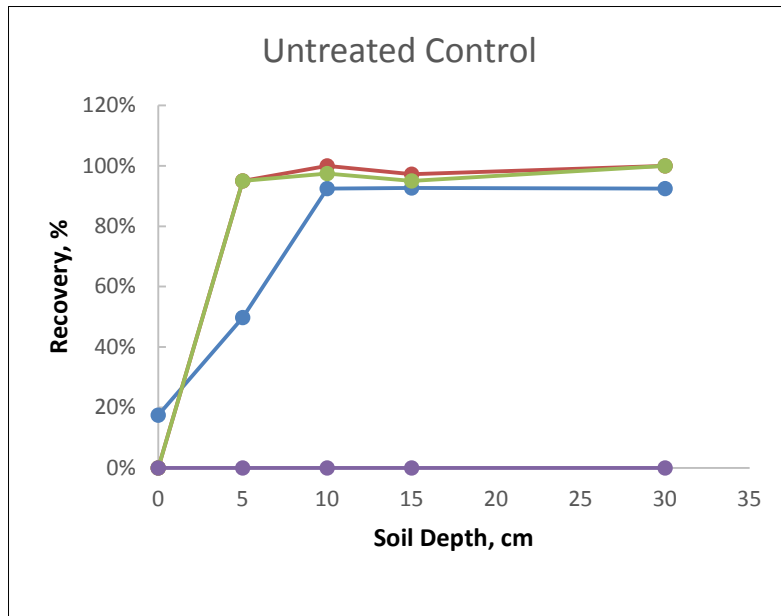
Pots and media can also be solarized



Soil solarization/steaming experiment at WSU Puyallup, July-August 2014



Recovery of *Phytophthora* spp. from various soil treatments – preliminary data



- P. plurivora*
- P. gonapodyides*
- P. taxon PgChlamydo*
- Noninoculated control

Steam treatment of contaminated soil

Soil should be at 50 C (125 F) for 50 minutes to kill pathogens

Moisture content and soil composition/bulk density will affect time needed to reach target temperature

Most of the inoculum will be at <5 cm, but may percolate further in looser soils



Data is collected on soil temperature at 3 depths during steaming



Sheet steaming and steam boiler. Photo: H.D. Siefert

Robot soil steamer

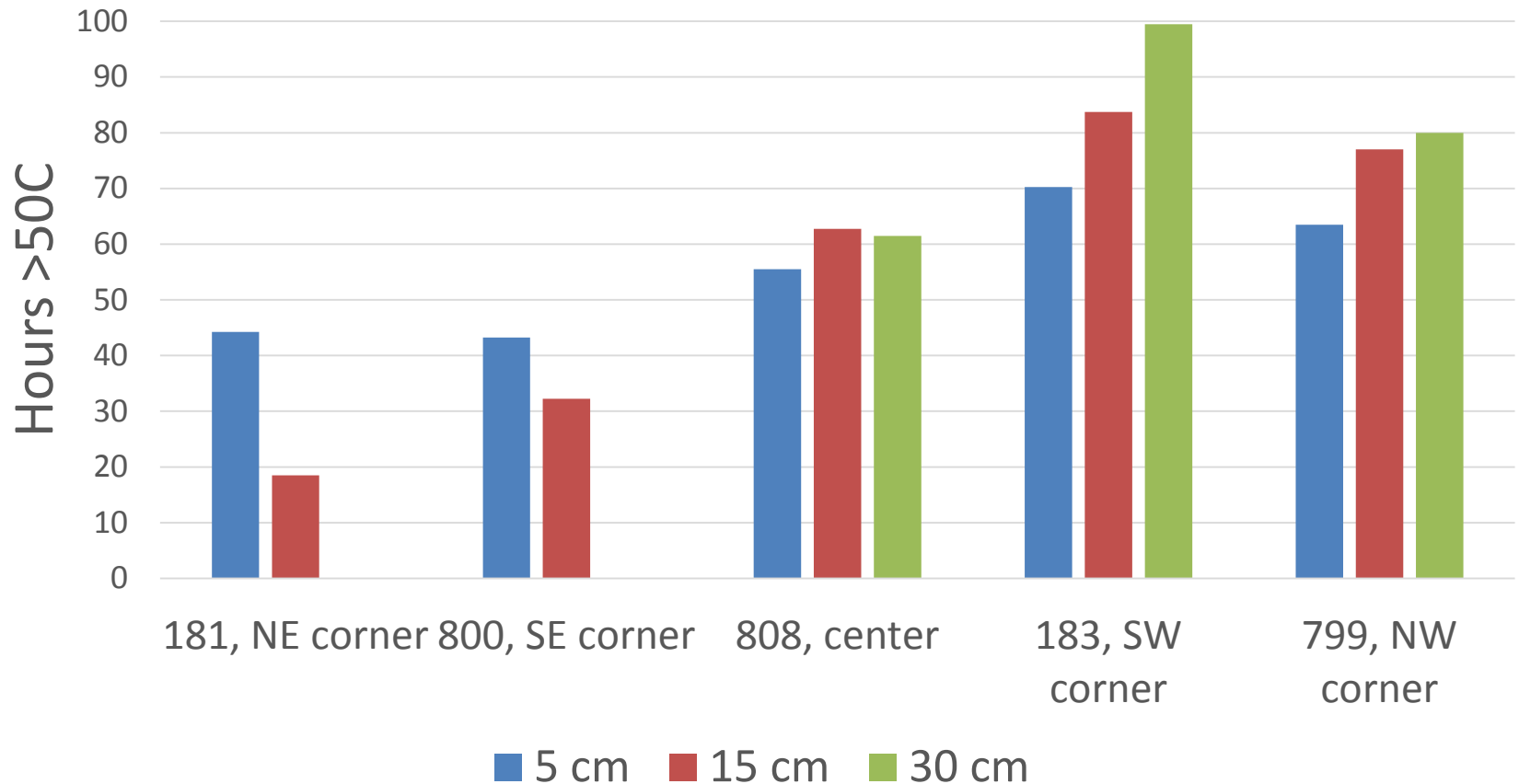


Information on this and other methods can be found at
<http://www.marten-barel-consultancy.com/steaming-methods/>



Steam treatment of *P. ramorum* positive soil at a nursery

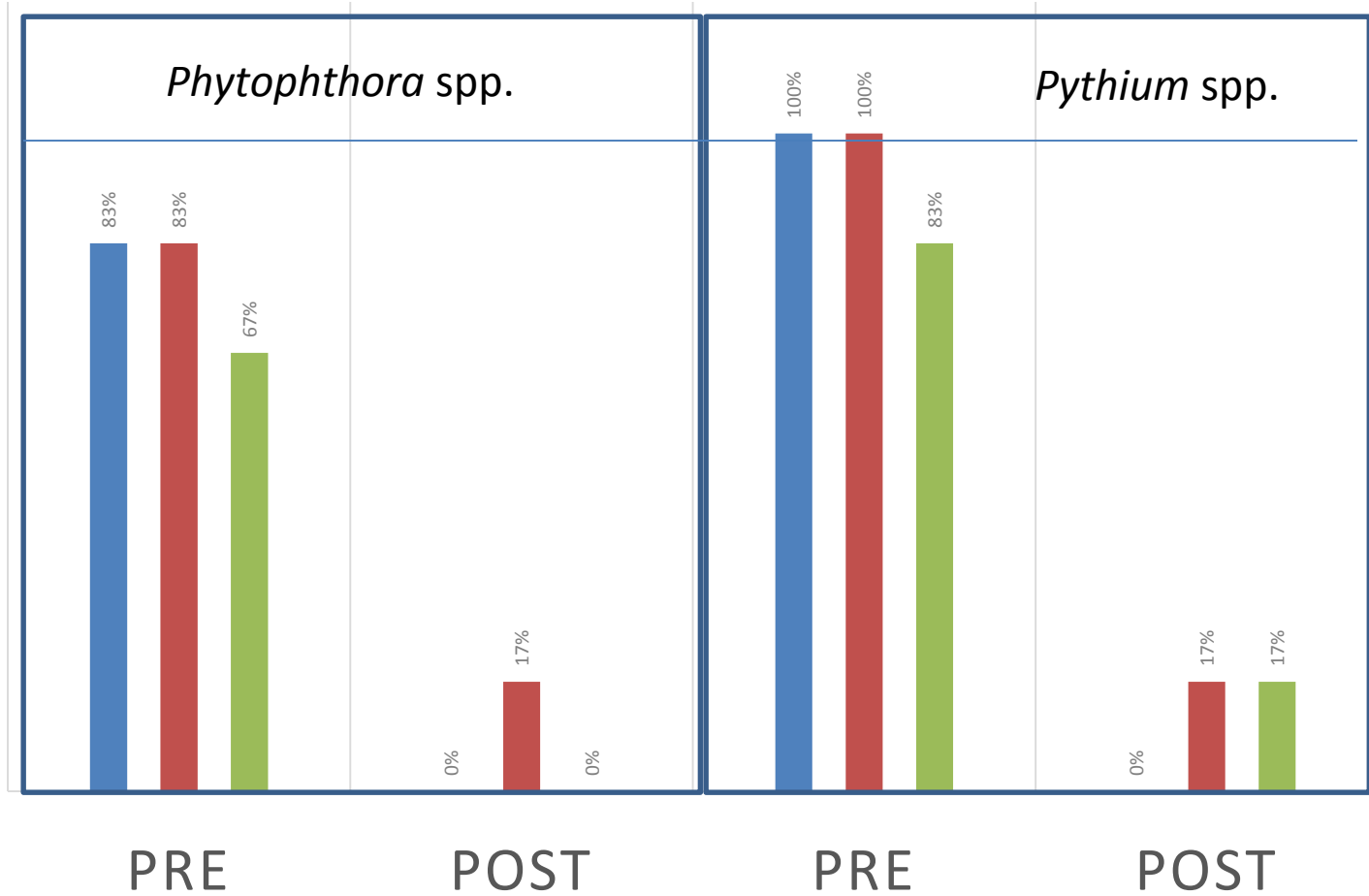
Area C1, plot size 15' x 20'



Survival

AREA C1

■ 0-10cm ■ 10-20cm ■ 20-30cm



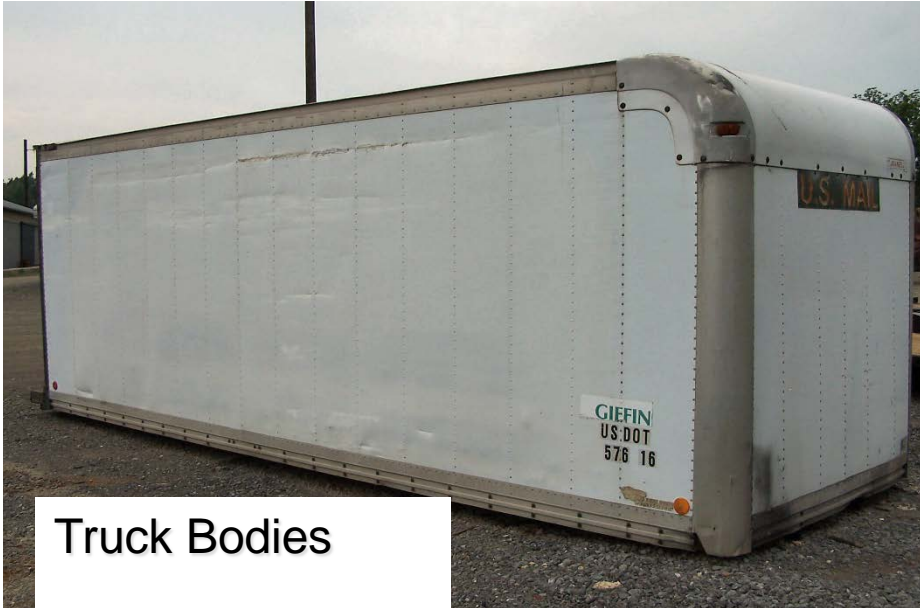
Steam Chamber for Pots

Compact, portable steamer





Steam Chamber Construction



Truck Bodies



Ocean Containers

Siebring steam cabinet

stainless steel cabinet accommodates pallets of pots



4. Which of these methods do you use to treat soil and/or potting media at your nursery?

Steam
Solarization
Chemical fumigation
Biocontrol
None
N/A