How To: Clean NMR Tubes

WILMAD NMR tubes are not 'analytically clean' when delivered to you. So if your NMR samples require scrupulously clean glass, follow the procedures below for Difficult Cleaning Problems to assure your sample purity is never jeopardized. Since NMR tubes are formed over a metal mandrel and certain organic lubricants are used, these cleaning steps will assure that any trace organic or inorganic residues from these procedures is removed.

Improper cleaning can damage NMR tubes and reduce your apparent spectrometer performance. You should never use a brush or other abrasive materials to clean NMR tubes. Scratches on the inside surface of the tube allow a portion of the sample to extend beyond the perfect cylinder defined by the NMR tube. Because the portion of your sample which fills a scratch on the inner surface of a tube experiences a different magnetic field than the rest of the sample, lines will broaden and resolution will deteriorate when you use scratched tubes.

Simple Cleaning of NMR Tubes

Cleaning your NMR tubes can be as simple as rinsing the tube with water or an organic solvent one at a time.

A final rinse with Acetone is frequently used to remove the last organic contents from the tube. When your sample is to be dissolved in water or D$_2$O, a final rinse with distilled water is usually adequate. You may want to take steps to remove traces of water from the surface of the tube outlined below.

Difficult Cleaning Problems

Tubes left with samples in them for a period of time frequently present a more challenging cleaning problem. Sample degradation or precipitation can cause material to adhere to the inner walls of the tube. Rinsing the tube doesn't always remove this adhered material. To remove adhered material use a strong mineral acid$^1$ such as concentrated or, in severe cases, fuming Nitric Acid soaks of 1-3 days, as needed. Nitric Acid can oxidize many organic chemicals and dissolves most inorganic materials, as well.

Copious rinsing of NMR tubes washed in acids is required to assure removal of residual acids. A final rinse with distilled water or Acetone is also appropriate.

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$^1$ Chromic Acid is not recommended, since residual Chromium can often adversely affect NMR experiments. Chromic Acid, while a stronger oxidizer, can leave paramagnetic Chromium VI behind, which can be removed only with repeated soaks with Nitric Acid.
Tubes which contained polymeric samples can be even more difficult to clean. When the polymers are natural products, like proteins and polysaccharides, strong acid soaks will usually be sufficient. However, when dealing with synthetic polymers, the challenge is more severe, since many polymers are inert to acids or insoluble in organic solvents by design. Although polymers may not readily dissolve in solvents, it may be possible to soften them by soaking the tubes in a solvent that swells the polymer. Then a pipe cleaner might be sufficient to remove the softened material. It may take some experimentation to find the solvent combination that works best with your polymer system.

Agitation in an Ultrasonic bath with an appropriate solvent can also help dislodge stubborn sample residues. However, you should take precautions to assure that NMR tubes do not touch, since contact and vibrations can fracture delicate thin wall tubes. WILMAD offers a special tube rack for use in its Ultrasonic bath that prevents such destructive contact between tubes.

**Removing Water from NMR Tubes**

Drying tubes at elevated temperatures can reshape and ruin NMR tubes. If you dry tubes in an oven, it is absolutely necessary to place tubes on a perfectly flat tray at 125° C for only 30 minutes. Better is the use of a vacuum oven that will remove water at lower temperatures. In a flat position, tubes that do reshape could be out-of-round and may not fit the spinner turbine as well. But they'll not affect the spectrometer probe adversely. Tubes placed in an oven in a beaker, flask, or tube rack can bend. Bent tubes may still fit the spinner, but can cause serious damage to the NMR probe, a costly repair with many probes.

But even drying at high temperatures doesn't remove water chemisorbed to the surface of the tube. Thus, the preferred method of water removal is chemical, not physical, treatment. In most cases, it is the protic content of water that must be avoided. So it is recommended to exchange the protons of chemisorbed water with a deuterated solvent such as D$_2$O prior to a short drying period in the oven. A bottle of D$_2$O that isn't being used any longer is perfect for this purpose.

When water chemically degrades your samples, then removal of water is essential. Here, reaction of the water with a hydride solution can be used, with caution. After rinsing the hydride solution, a final rinse with very dry Acetone can be used to remove rinse solvent prior to oven drying. Cap tubes promptly to avoid absorption of moisture when removing dry tubes from the oven.